

Water-soluble Constituents from *Callicarpa pedunculata*

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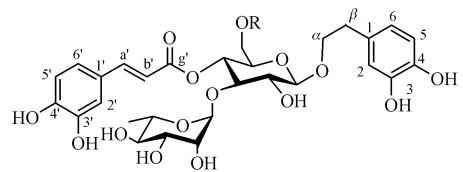
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ABSTRACT **AM:** To explore the water-soluble constituents from *Callicarpa pedunculata*. **METHOD:** The water-soluble constituents were isolated and purified by column chromatography and their structures were identified by spectroscopic analyses including 1D and 2D NMR data. **RESULTS:** Four caffeoyl phenylethanoid glycosides were purified from the water-soluble extracts of *Callicarpa pedunculata* R. Brown and were identified to be forsythoside B (1), arenarioside (2), acteoside (3) and isoacteoside (4) respectively. **CONCLUSION:** Compounds 1-4 were obtained from this plant for the first time.

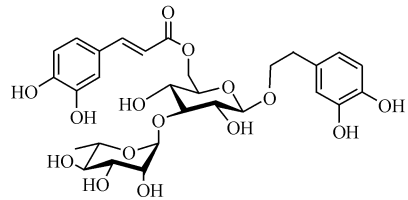
KEY WORDS *Callicarpa pedunculata*; Caffeoyl phenylethanoid glycosides; Forsythoside B; Arenarioside; Acteoside; Isoacteoside

CLC number R284.1 **Document code** A **Article ID** 16722365.1(2008)03:0176:20
doi: 10.3724/SP.J.1009.2008.00176

Callicarpa pedunculata R. Brown (Verbenaceae) is a small shrub widely distributed in southeast and southern China to the Philippines and has been used in traditional Chinese medicine for the treatment of hemostasis and as anti-inflammatory and antibacterial drugs^[1]. In previous papers, we reported the structures of four types of diterpenoids, flavones and triterpenoids^[2,3]. Our continuing phytochemical investigation on water-soluble constituents of this plant, four phenylpropanoids were isolated and their structures were elucidated as forsythoside B (1), arenarioside (2), acteoside (3) and isoacteoside (4) respectively.



- 1 R = β -D-Api
2 R = β -D-Xyl
3 R = H



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1 Apparatus and Material

Column chromatography (CC): Qingdao silica gel (200-300 mesh); macroporous resin D101 (Nankai University); Sephadex LH20 Pharmacia products. **TLC:** Qingdao precoated plates, silica GF₂₅₄ plates. **NMR Spectra:** Bruker DRX2500 spectrometer with TMS as internal standard. **ESIMS:** ThermoFinnigan LCQ2 Advantage spectrometer.

2 Plant material

The aerial parts of *C. pedunculata* R. Brown were collected in Jiangxi Province, China and a voucher specimen was deposited in the herbarium of the Kunming Institute of Botany, Chinese Academy of Sciences (No. 0300993).

Received on: 2007-07-23

Foundation Item This project was supported by the National Natural Science Foundation for Outstanding Youth (No. 30325044) to SHEN YueMao.

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3 Extraction and Isolation

The air-dried aerial parts of this plant (150 g) were extracted five times with boiling water. The aqueous solution was concentrated under vacuum at 45°C by coevaporation with *n*-butanol. The concentrate solution (ca. 400 mL) was subjected to column chromatography over macroporous resin D101 (250 g) eluted with water (4 L), 20% ethanol (V/V, 4 L) and ethanol (3 L), respectively. The 20% ethanol eluate was collected with 500 mL per fraction. The fractions 228 were combined according to the TLC results (TLC plates were precoated with silica gel F₂₅₄, and developed with *n*-BuOH:H₂O:FA:8:1:1, V/V/V) to afford 113 g extract after the removal of solvents under vacuum. This extract was subjected to column chromatography over reversed-phase C₁₈ silica

gel (145 g) eluted with gradient water-methanol (H₂O 20:18 L; 30% MeOH 22:15 L; 60% MeOH 21:10 L; MeOH) and collected as 500 mL/fraction. The 60% MeOH eluate were combined according to the TLC results and further subjected to column chromatography over Sephadex LH 20 (120 g) eluted with 85% MeOH (V/V) to yield 1 (8 mg yield cal 010053%) and 2 (27 mg yield cal 01018%)¹

In order to prepare more amounts of compounds 1 and 2, larger scale extraction with more plant materials was carried out in pharmaceutical factory. The isolation procedure was the same as described above.

However, compounds 3 and 4 were obtained as a mixture (the ratio of 3 to 4 was estimated to be 1:3 based on the integrations of proton NMR signals yield cal 1/10 of 1) besides 1 and 2 this time.

4 Structure Elucidation

The structures of 1-4 were elucidated by comparison of the spectral data (¹H NMR and ¹³C NMR, Tables 1 and 2) with those reported data and 2D NMR spectrum to be forsythoside B^[5], arenarinoside^[6], acteoside^[7,8] and isoacteoside^[9], respectively.

Table 1 ¹H NMR data of compounds 1-4^a (500 MHz in MeOD)

Position of aglycone	1 ¹ H	2 ¹ H ^b	3 ¹ H	4 ¹ H ^b
1	-	-	-	-
2	61.70 (d, 21.0)	61.61 (d, 21.0)	61.71 (d, 21.0)	61.68 (d, 21.0)
3	-	-	-	-
4	-	-	-	-
5	61.70 (d, 81.0)	61.73 (d, 81.0)	61.73 (d, 81.0)	61.65 (d, 81.0)
6	61.58 (dd, 21.0, 81.0)	61.47 (dd, 21.0, 81.0)	61.57 (dd, 21.0, 81.0)	61.53 (dd, 21.0, 81.0)
A	31.71 (m), 41.08 (m)	31.63 (m); 31.90 (m)	31.70 (m); 41.02 (m)	31.70 (m); 41.02 (m)
B	21.82 (d, 21.7, 71.0)	21.68 (m, 2H)	21.77 (m, 2H)	21.77 (m, 2H)
caffeoyl moiety				
1c	-	-	-	-
2c	71.07 (d, 21.0)	61.96 (d, 21.0)	71.07 (d, 21.0)	71.04 (d, 21.0)
3c	-	-	-	-
4c	-	-	-	-
5c	61.79 (d, 81.0)	61.69 (d, 81.0)	61.80 (d, 81.0)	61.78 (d, 81.0)
6c	61.97 (dd, 21.0, 81.0)	61.86 (dd, 21.0, 81.0)	61.96 (dd, 21.0, 81.0)	61.88 (dd, 21.0, 81.0)
Ac	61.29 (d, 151.9)	61.19 (d, 151.8)	61.29 (d, 151.8)	61.29 (d, 151.8)
Bc	71.61 (d, 151.9)	71.51 (d, 151.8)	71.61 (d, 151.8)	71.56 (d, 151.8)
Cc	-	-	-	-
glucose				
1	41.37 (d, 71.7)	41.27 (d, 71.9)	41.38 (d, 71.9)	41.34 (d, 71.8)
2		31.31 (t, 81.2)	31.53 ^c	31.36 ^c
3		31.71 (m)	^c	31.60 (m)
4	41.97	41.90 (t, 91.5)	41.90 (m)	^c
5		31.63 (m)	31.53 ^c	31.36 ^c
6		31.49 (m); 31.77 (m)	31.56 ^c ; 31.70 ^c	41.38 (m); 41.51 (m)
rhamnose				
1	51.17 (d, 11.3)	51.07 (br s)	51.19 (br s)	51.19 (br s)
2		31.83 (dd, 11.8, 31.0)	^c	^c
3		31.49 (m)	^c	^c
4		31.21 (t, 91.6)	31.32 ^c	31.45 ^c
5		31.49 (m)	^c	^c
6	11.09 (d, 61.1)	01.98 (d, 61.2, 3H)	11.10 (d, 61.2, 3H)	11.27 (d, 61.2, 3H)
apiose		xy1		
1	41.91 (d, 21.2)	41.14 (d, 71.5)		
2	31.68 (d, 21.2)	31.13 (dd, 71.6, 81.9)		
3	-	31.23 (t, 81.9)		
4	31.60/31.56	31.37 (m)		
5	31.31	31.06 (t, 101.9); 31.74 (m)		

a ¹H NMR spectra were obtained at 500 MHz and recorded in MeOD at room temperature.

b Coupling constants are presented in Hertz. Unless otherwise indicated, all proton signals integrate to 1H.

c The proton signals overlapped severely.

Tabele 2 ¹³C NMR data of compounds 1-4 (125 MHz in Me₂CO)

Position of aglycone	1 ¹³ C	2 ¹³ C	3 ¹³ C	4 ¹³ C
1	131.4	131.4	131.4	131.3
2	115.3	117.1	117.1	117.1
3	146.0	145.9	145.9	145.9
4	144.5	144.5	144.4	144.4
5	116.3	116.3	116.4	116.4
6	121.3	121.3	121.3	121.3
A	72.2	72.3	72.3	72.2
B	36.5	36.4	36.3	36.4
caffeoyl moiety				
1c	127.6	127.5	127.6	127.6
2c	114.7	115.3	115.6	115.3
3c	146.7	146.6	146.5	146.5
4c	149.6	149.7	149.5	149.4
5c	116.5	116.5	116.5	116.5
6c	123.2	123.3	123.2	123.2
Ac	117.1	114.5	114.6	114.7
Bc	148.0	148.2	148.1	147.2
Cc	168.2	168.4	168.3	169.7
glucose				
1	104.1	104.0	103.9	104.1
2	81.6	75.9	76.0	75.5
3	76.0	81.6	81.7	83.8
4	70.4	70.3	70.4	70.2
5	74.4	74.6	75.7	75.1
6	68.3	69.2	62.2	64.5
rhamnose				
1	103.0	102.9	102.9	102.5
2	72.2	72.2	72.3	72.1
3	72.0	71.9	71.9	72.1
4	73.7	73.6	73.7	73.8
5	70.8	70.3	70.3	69.9
6	18.4	18.4	18.4	17.8
apiose		xyl		
1	110.9	105.1		
2	78.0	74.7		
3	80.6	77.4		
4	75.0	71.0		
5	65.6	66.7		

5 Discussion

Caffeoyl phenylethanoid glycosides have been found in the families of Scrophulariaceae, Oleeaceae,

Plantaginaceae, Lamnaceae and Orobanchaceae, and the genera Clerodendron, Lantana, Petrea and Verbenaceae^[10]. Till 2005, Koo et al^[11] isolated phenylethanoid glycosides from *Calliandra dichotoma* and reported their neuroprotective activities and anti-amnesic activity of acteoside^[12]. Our work is the first report about the isolation of caffeoyl phenylethanoid glycosides from the plant *Calliandra dichotoma*.

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紫珠的水溶性成分

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摘要 目的: 研究中药紫珠的水溶性成分。方法: 运用多种色谱技术进行分离纯化, 运用光谱技术鉴定化合物结构。结果: 从紫珠的水溶性提取物中分离到 4 个苯丙素苷类化合物, 分别鉴定为连翘苷 (forsythoside B 1), arenarionolide (2), 类叶升麻苷 (acteoside 3) 和异类叶升麻苷 (isoacteoside 4)。结论: 这 4 个化合物都是首次从该植物中分离得到。

关键词 紫珠; 苯丙素苷; Forsythoside B; Arenarionolide; Acteoside; Isoacteoside

基金项目 国家杰出青年基金 (No. 30325044)