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Claviconol, a new metabolite, from the mycelia culture of *Claviconora pyxidata*

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Abstract: **Aim** To explore the new bioactive metabolites from the mycelia culture of the edible mushroom *Claviconora pyxidata*. **Methods** The constituents were isolated and purified by column chromatography, and their structures were identified by spectroscopic analyses, including 1D- and 2D-NMR data, and single-crystal X-ray diffraction. **Results** A new natural product, named claviconol (**1**) along with adenosine (**2**), were obtained. **Conclusion** Compound **1** is a new compound and stereochemistry of **2** was confirmed by a single-crystal X-ray diffraction.

Key words: *Claviconora pyxidata*; claviconol; adenosine; single-crystal X-ray diffraction

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珊瑚菌菌体培养物的新天然产物

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摘要: 目的 对珊瑚菌 (*Claviconora pyxidata*) 菌体培养物的活性成分进行分离。方法 用多种色谱技术对化合物进行分离纯化, 并用光谱技术和单晶 X 射线衍射技术鉴定化合物的结构。结果 从中分离到 2 个化合物: claviconol (**1**)、腺苷 (**2**)。结论 化合物 **1** 为新化合物, 并通过单晶 X 射线衍射技术确定了 **2** 的立体构型。

关键词: 珊瑚菌; claviconol; 腺苷; 单晶 X 射线衍射

Claviconora pyxidata is a wild mushroom which was widely used for curing gastric pain, dyspepsia, gout and heat-toxicity in China. Some compounds have been isolated from *Claviconora pyxidata*, such as claviconic acid which was a novel inhibitor of reverse transcriptases^[1]. *Claviconora pyxi-*

data YB2005 was isolated from the wild fruit body in Jilin Province and identified by ITS methods. In exploring the new bioactive metabolites from this medicinal mushroom, a new metabolite, claviconol (**1**) along with adenosine (**2**) were obtained. Their structures were elucidated by the analysis of NMR

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data (^1H , ^{13}C -NMR, DEPT, HSQC, HMBC, ^1H - ^1H COSY and NOESY). The relative stereochemistry of **2** was confirmed by a single-crystal X-ray diffraction

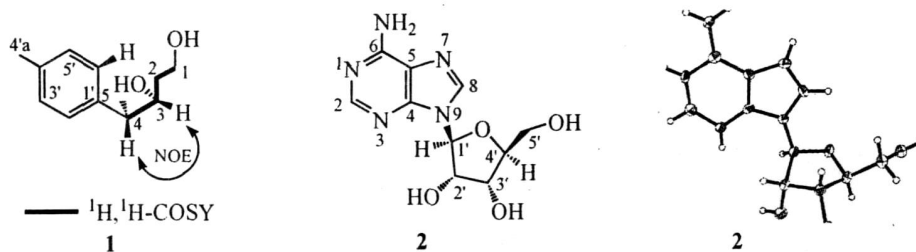


Figure 1 The structures of **1** and **2**

Clavicornol (**1**) was obtained as a colorless oil, [$\alpha_D^{20} + 4.56$ (c 0.29, CHCl_3)]. The molecular formula was determined as $\text{C}_{12}\text{H}_{18}\text{O}_2$ according to the HR ESI-Q-TOF MS which showed the obvious quasi-molecular ion at m/z 217.0933 [$\text{M} + \text{Na}$] $^+$. IR absorption implied the presence of hydroxyl (3353 cm^{-1}), methyl (2924 cm^{-1}) and aromatic (1593 cm^{-1}) groups. ^1H -NMR spectra of **1** (Table 1) showed the presence of four aromatic protons [at 7.09 (d, 2H, $J = 7.8\text{ Hz}$); 7.17 (d, 2H, $J = 7.8\text{ Hz}$)] and indicated the 1, 4-bisubstitutions of aromatic moiety. The presence of aromatic moiety was further supported by the ^{13}C -NMR resonances assigned to six aromatic carbons (Table 1). HMBC correlation from the methyl protons at 2.28 (H-4a) to C-3, C-4 and C-5 indicated the linkage of the methyl at C-4. The cross peaks between H-5 and H-4, H-4 and H-3, H-3 and H-2b, and H-1 and H-2a (2b) can be observed in the ^1H - ^1H -COSY spectrum and allowed to establish the fragment of CH_3 (5)-CH (4)-CH (3)- CH_2 (2)- CH_2 (1) (Figure 1). The key HMBC correlations from H-5 to C-4, C-3 and C-1, and H-4 to C-2, C-3, C-1 and C-2 (6), and H-1

to C-3 confirmed the connections of the fragment and allowed the linkage of the fragment with C-1. The hydroxyl groups were determined at C-1 and C-3 according to the downshift of the carbons at 61.0 and 74.9, respectively. The relative stereochemistry of **1** was determined by the NOE correlations between H-4 and H-3.

Adenosine (**2**) was elucidated by analysis of NMR data (^1H , ^{13}C NMR, HSQC, HMBC) (Table 2). Compound **2** was an enantiomer of β -adenosine whose NMR data (^1H , ^{13}C) was reported by Ciuffreda^[21] and coworkers. The stereochemistry of **2** was established by single-crystal X-ray diffraction (CCDC number: 685692) (Figure 1). It was reported that some edible fungi contained the high level of nucleosides and nucleobases^[31]. Adenosine, one of nucleoside, possessed many kinds of bioactivities and have been used in clinic.

Compounds **1** - **2** showed no antibacterial (*Escherichia coli*, *Bacillus Subtilis* and *Staphylococcus aureus*) and anti-yeast (*Candida albicans*) activities at the concentration $100\text{ }\mu\text{g}\cdot\text{mL}^{-1}$ using a similar MIC method and exhibited low cytotoxicities against HeLa cells by MTT assay.

Table 1 ^1H -NMR and ^{13}C -NMR data for clavicornol (**1**) in $(\text{CD}_3)_2\text{CO}$

| Position | ^1H (600 MHz) | ^{13}C (150 MHz) | HMBC (600 MHz) |
|----------|------------------------------------|---------------------------|-----------------------------|
| 1 | 3.70 (d, 1H, $J = 5.4\text{ Hz}$) | 61.0 (t) | C-3 |
| 2a | 1.64 (m, 1H) | 37.3 (t) | C-1, C-3 |
| 2b | 1.48 (m, 1H) | | |
| 3 | 3.91 (m, 1H) | 74.9 (d) | |
| 4 | 2.78 (m, 1H) | 46.4 (d) | C-2, C-3, C-5, C-1, C-2 (6) |
| 5 | 1.27 (d, 3H, $J = 7.2\text{ Hz}$) | 17.7 (q) | C-3, C-4, C-1 |
| 1 | | 142.1 (s) | |
| 2 | 7.09 (d, 1H, $J = 7.8\text{ Hz}$) | 129.4 (d) | C-4, C-4, C-3 |
| 3 | 7.17 (d, 1H, $J = 7.8\text{ Hz}$) | 129.2 (d) | C-4, C-4 a, C-2 |

(to be continued)

Continued Table 1

| Position | ^1H (600 MHz) | ^{13}C (150 MHz) | HMBC (600 MHz) |
|----------|----------------------------|---------------------------|-----------------|
| 4 | | 135.9 (s) | |
| 4 a | 2.28 (s, 3H) | 21.0 (q) | C-4, C-3 (5) |
| 5 | 7.17 (d, 1H, $J = 7.8$ Hz) | 129.2 (d) | C-4, C-4 a, C-6 |
| 6 | 7.09 (d, 1H, $J = 7.8$ Hz) | 129.4 (d) | C-4, C-4, C-5 |
| OH | 3.64 (overlap, 1H) | | |
| OH | 3.64 (overlap, 1H) | | |

Table 2 ^1H -NMR and ^{13}C -NMR data for adenosine (2) in $\text{DMSO}-d_6$

| Position | ^1H (600 MHz) | ^{13}C (150 MHz) | HMBC (600 MHz) |
|---------------|----------------------------|---------------------------|----------------|
| 2 | 8.13 (s, 1H) | 152.4 (d) | C-4, C-6 |
| 4 | | 149.0 (s) | |
| 5 | | 119.3 (s) | |
| 6 | | 156.1 (s) | |
| 8 | 8.34 (s, 1H) | 139.9 (d) | C-4, C-5, C-1 |
| 1 | 5.88 (d, 1H, $J = 6.1$ Hz) | 87.9 (d) | C-4, C-8, C-2 |
| 2 | 4.61 (d, 1H, $J = 6.1$ Hz) | 73.4 (d) | C-1, C-4 |
| 3 | 4.14 (d, 1H, $J = 1.9$ Hz) | 70.6 (d) | C-1, C-5 |
| 4 | 3.96 (d, 1H, $J = 1.9$ Hz) | 85.9 (d) | C-3 |
| 5 a | 3.68 (m, 1H) | 61.6 (t) | C-3, C-4 |
| 5 b | 3.56 (m, 1H) | | |
| NH_2 | 7.36 (s, 2H) | | C-6 |
| 2'-OH | 5.47 (overlap, 1H) | | C-1, C-2, C-3 |
| 3'-OH | 5.21 (d, 1H, $J = 3.9$ Hz) | | C-2, C-3, C-4 |
| 5'-OH | 5.46 (overlap, 1H) | | C-4, C-5 |

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